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Dehydrative glycosylation of tri-O-benzylated 1-hydroxyribofuranose catalyzed by a copper(II) complex

Takeyuki Suzuki, Shoko Watanabe, Taichiro Yamada and Kunio Hiroi*

Department of Synthetic Organic Chemistry, Tohoku Pharmaceutical University, 4-4-1 Komatsushima, Aoba-Ku, Sendai 981-8558, Japan

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Abstract—A phosphine/Cu(II) complex catalyzes the dehydrative glycosylation of tri-O-benzylated 1-hydroxyribofuranose to give the ribofuranoside with high stereoselectivity. © 2003 Elsevier Science Ltd. All rights reserved.

The development of efficient stereoselective glycosylations is one of the most important subjects in carbohydrate chemistry.1 Although the direct glycosylation of 1-hydroxy sugars is highly desirable,² there are only a few examples of catalytic dehydrative glycosylations. Inanaga reported that methoxyacetic acid/Yb(OTf)₃ (10 mol%) promotes the reaction of 1-hydroxy sugars with alcohols to afford the glycosides in 72-99% yield with moderate to excellent selectivities.3 Mukaiyama developed some elegant methods for the stereoselective synthesis of both 1,2-cis- and trans-ribofuranosides from 1-hydroxyribofuranoses and alcohols or trimethylsilyl ethers using Ph₂Sn=S (20 mol%)/AgClO₄,⁴ Lawesson's reagent (10 mol%)/AgClO₄, 4 M(OTf)n (1 mol%)/hexamethyldisiloxane (M = Sn, La, and Yb), $TrB(C_6F_5)_4$ (3 mol%).⁶ We report a new protocol using

a phosphine Cu(II) catalyst that gives the ribofuranosides with reasonable selectivity (Scheme 1).

When a mixture of 2,3,5-tri-O-benzyl-D-ribofuranose (1)⁷ and cyclohexanol (2a) and a catalyst system prepared from CuCl₂, bis(diphenylphosphino)ferrocene (dppf), and AgClO₄ (1:1:2 mol ratio) in CHCl₃ in the presence of CaSO₄ as a dehydrating agent (1:2a:Cu:Ca=100:120:5:1000 mol ratio) was allowed to stand at room temperature for 21 h, cyclohexyl 2,3,5-tri-O-benzyl-D-ribofuranoside (3a)^{2c} was obtained in 95% yield (α : β =5:95). No reaction occurred without CuCl₂ or AgClO₄ under otherwise identical conditions. The use of CuCl instead of CuCl₂ also afforded no reaction. Changing the Cu/AgClO₄ ratio affected the reactivity. The catalyst prepared with a Cu/dppf/

BnO OH + ROH (R'SH) Cast.
$$\frac{\text{cat. CuCl}_2}{\text{cat. dppf}}$$
 $\frac{\text{cat. CuCl}_2}{\text{cat. dppf}}$ $\frac{\text{cat. CuCl}_2}{\text{cat. dppf}}$ $\frac{\text{cat. AgClO}_4}{\text{CaSO}_4, \text{CHCl}_3}$ $\frac{\text{BnO}}{\text{BnO}}$ $\frac{\text{OR}}{\text{(SR')}}$ $\frac{\text{BnO}}{\text{CaSO}_4, \text{CHCl}_3}$ $\frac{\text{BnO}}{\text{CaSO}_4, \text{CHCl}_3}$ $\frac{\text{BnO}}{\text{BnO}}$ $\frac{\text{CasO}_4}{\text{CaSO}_4, \text{CHCl}_3}$ $\frac{\text{BnO}}{\text{BnO}}$ $\frac{\text{CasO}_4}{\text{CaSO}_4, \text{CHCl}_3}$ $\frac{\text{BnO}}{\text{BnO}}$ $\frac{\text{CasO}_4}{\text{CasO}_4, \text{CHCl}_3}$ $\frac{\text{CasO}_4}{\text{CasO}_4, \text{CasO}_4, \text{CasO}_4}$ $\frac{\text{CasO}_4}{\text{CasO}_4, \text{CasO}_4, \text{CasO}_4$

Scheme 1. Dehydrative glycosylation of 2,3,5-tri-*O*-benzyl-D-ribofuranose 1.

^{*} Corresponding author. Tel.: +81 22 234 4181 (ext. 2531); fax: +81 22 275 2013; e-mail: khiroi@tohoku-pharm.ac.jp

Table 1. Dehydrative glycosylation of 1 catalyzed by a Cu(II) complex^a

Entry	Acceptor	Additive	Time (h)	Ribofuranoside		
				No.	Yield ^b (%)	α/β^c
1	2a	None	21	3a	95	5:95
2	2a	LiClO ₄	7	3a	90	96:4
3	2 b	None	22	3b	85	10:90
4	2b	LiClO ₄	3	3b	87	88:12
5	2c	None	22	3c	80	15:85
6	2c	LiClO ₄	6	3c	97	93:7
7	2d	None	16	3d	77	19:81
8	2d	LiClO ₄	5	3d	92	92:8
9	2e	None	19	3e	77	25:75
10	2e	LiClO ₄	7	3e	82	98:2
11	2f	None	19	3f	68	93:7 ^d
12	2f	LiClO ₄	19	3f	89	71:29 ^d

^a The reaction was conducted at room temperature using 0.100 mmol of 1, 0.120 mmol of 2, 1.00 mmol of CaSO₄, and 0.150 mmol of additive in CHCl₃ (0.02 M) containing 5 mol% of the catalyst.

AgClO₄ (1:1:1 mol ratio) gave a lower yield of 39%. These results suggest that Cu(ClO₄)₂ is an active catalyst. In fact, Cu(ClO₄)₂·6H₂O could be used in place of CuCl2-AgClO4. Other silver salts such as AgOTf, AgBF₄, AgPF₆, and AgSbF₆ gave less satisfactory results. The activity of CuCl₂-AgClO₄ was enhanced by the addition of 1 equiv. of dppf at the early stage of the reaction. The yield after 1 h was increased from 13 to 87%. Although other phosphine ligands such as dppp and dppb showed similar reactivities and selectivities, the addition of bipyridine completely retarded the reaction. It is noteworthy that reversed stereoselectivity was obtained by the addition of 1.5 equiv. of LiClO₄ to the catalyst system.9 Thus, the addition of LiClO4 to the reaction mixture of 1 and 2a afforded 3a in 90% yield $(\alpha:\beta=96:4).$

Table 1 shows some examples of the dehydrative glycosylation of 1 with various glycosyl acceptors using the phosphine/Cu catalyst.¹⁰ The reaction of i-C₃H₇OH 2b gave the ribofuranoside 3b in 85% yield (α: β =10:90, entry 3). Primary alcohols 2c, 2d and 2e can be used as glycosyl acceptors to give 3c, 3d^{2c} and 3e,¹¹ although the extent of β -selectivity is less satisfactory (entries 5, 7 and 9). However, The reaction of 2e in the presence of LiClO₄ gave 3e with the highest α -selectivity (entry 10). Interestingly, the reaction of thiophenol afforded the corresponding α -ribofuranoside 3f¹² preferentially with or without LiClO₄ (entries 11 and 12).

In summary, a Cu(II)-catalyzed dehydrative glycosylation of tri-O-benzylated 1-hydroxyribofuranose was developed for the first time. The properties of the Cu(II) catalysts should be readily modifiable to afford more efficient catalysts, because various phosphine ligands are available. Our results should provide a basis for designing more efficient catalyst systems.

References

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^b Isolated yield.

^c HPLC analysis using a Tosoh TSKgel Silica-60 column unless otherwise specified.

^d Determined by ¹H NMR analysis.

- 10. General procedure for glycosylation of 1 using Cu(II) catalyst: To a stirred suspension of CuCl₂ (0.7 mg, 0.005 mmol), dppf (2.8 mg, 0.005 mmol), AgClO₄ (2.1 mg, 0.010 mmol) and CaSO₄ (136 mg, 1.0 mmol) in CHCl₃ (2 mL) was added a solution of 1 (42.1 mg, 0.1 mmol) and 2 (0.12 mmol) in CHCl₃ (3 mL) at room temperature. After the time shown in Table 1, the mixture was diluted with ether and the solution was passed through a Celite pad, and concentrated under reduced pressure. The resulting crude product was purified by flash chromatography on silica gel. Spectral data for **3b**. α -Anomer: $[\alpha]_D^{26}$ +57.1 (c 1.19 CHCl₃); ¹H NMR (270 MHz, CDCl₃) δ 1.22 (d, J=6.1 Hz, 3H), 1.29 (d, J=6.1 Hz, 3H), 3.36 (dd, J=10.5, 4.1 Hz, 1H), 3.45 (dd, J=10.6, 3.6 Hz, 1H),3.75 (dd, J=6.6, 4.1 Hz, 1H), 3.82 (dd, J=6.8, 4.0 Hz,1H), 3.96 (sep., J = 6.1 Hz, 1H), 4.23 (q, J = 3.8 Hz, 1H), 4.4-4.8 (m, 6H), 5.11 (d, J=4.0 Hz, 1H), 7.2-7.4 (m, 15H); 13 C NMR (100 MHz, CDCl₃) δ 21.76, 23.65, 69.97, 70.00, 72.17, 72.35, 73.39, 75.41, 77.41, 81.02, 99.68, 127.48, 127.58, 127.62, 127.68, 127.98, 128.05, 128.19, 128.30, 128.32, 138.05, 138.13, 138.55; IR (neat) 3088, 3063, 3030, 2971, 2922, 2866, 1605, 1497, 1454, 1366, 1331, 1258, 1208, 1113, 1028, 737, 698; MS (EI) m/z 91 (Bn, base peak), 462 (M+); HRMS (EI); calcd for $C_{29}H_{34}O_5$ 462.2406 found 462.2413. β -Anomer: $[\alpha]_D^{26} + 2.8$ (c 1.06 CHCl₃); ¹H NMR (270 MHz, CDCl₃) δ 1.09 (d, J=6.1 Hz, 3H), 1.10 (d, J=6.1 Hz, 3H), 3.52 (dd, J=10.5, 6.0 Hz, 1H), 3.60 (dd, J=10.5, 4.1 Hz, 1H), 3.82 (bd, J=4.8 Hz, 1H), 3.88 (sep., J=6.1 Hz, 1H), 3.99–4.03 (m, 1H), 4.3–4.4 (m, 1H), 4.4–4.7 (m, 6H), 5.11 (s, 1H), 7.2–7.4 (m, 15H); 13 C NMR (100 MHz, CDCl₃) δ 21.33, 23.42, 69.26, 71.74, 72.39(2C), 73.16, 78.81, 80.23, 80.31, 103.33, 127.50, 127.67, 127.70, 127.76, 127.81, 127.96, 128.29, 128.34, 128.39, 137.99, 138.02, 138.35; IR (neat) 3088, 3063, 3030, 2971, 2926, 2863, 1605, 1497, 1454,
- 1368, 1308, 1262, 1208, 1124, 1096, 1028, 737, 698; MS (EI) m/z 91 (Bn, base peak), 462 (M⁺); HRMS (EI); calcd for $C_{29}H_{34}O_5$ 462.2406, found 462.2419. **3c**. α -Anomer: $[\alpha]_{D}^{22}$ +81.4 (c 1.57, CHCl₃); ¹H NMR (270 MHz, CDCl₃) δ 0.88 (t, J= 6.9 Hz, 3H), 1.26 (s, 12H), 3.37 (dd, J = 10.4, 4.1 Hz, 1H), 3.45 (dd, J = 10.5, 3.6 Hz, 1H), 3.52 (dt, J=9.9, 6.8 Hz, 1H), 3.7-3.85 (m, 3H), 4.23 (q, J=4.0)Hz, 1H), 4.2-4.75 (m, 6H), 5.00 (d, J=4.1 Hz, 1H), 7.1–7.4 (m, 15H); 13 C NMR (68 MHz, CDCl₃) δ 14.21, 22.76, 26.23, 29.39, 29.50, 29.72, 31.92, 68.48, 70.02, 72.21, 72.45, 73.40, 75.36, 81.34, 101.30, 127.43, 127.53, 127.60, 127.87, 127.99, 128.11, 128.19, 128.23, 137.89, 137.93, 138.31; IR (neat) 3088, 3063, 3030, 2926, 2857, 1607, 1497, 1454, 1360, 1331, 1258, 1208, 1115, 1080, 1044, 1028, 737, 698; MS (EI) m/z 91 (Bn, base peak), 532 (M⁺); HRMS (EI); calcd for C₃₄H₄₄O₅ 532.3189, found 532.3197. β-Anomer: $[\alpha]_D^{24}$ –21.6 (*c* 1.00, CHCl₃); ¹H NMR (270 MHz, CDCl₃) δ 0.88 (t, J=6.4 Hz, 3H), 1.25 (s, 12H), 3.32 (dt, J=9.6, 6.8 Hz, 1H), 3.51 (dd, J = 10.6, 5.9 Hz, 1H), 3.61 (dd, J = 10.6, 4.0 Hz, 1H), 3.66 (dt, J=9.6, 6.8 Hz, 1H), 3.86 (dd, J=4.8, 1.2 Hz, 1H),4.01 (dd, J=6.9, 4.6 Hz, 1H), 4.33 (m, 1H), 4.4-4.9 (m, 6H), 5.00 (d, J=1.2 Hz, 1H), 7.2–7.4 (m, 15H); ¹³C NMR (68 MHz, CDCl₃) δ 14.20, 22.74, 26.17, 29.32, 29.45, 29.57, 31.90, 68.00, 71.57, 72.27, 72.38, 73.14, 78.69, 79.86, 80.32, 105.25, 127.37, 127.49, 127.60, 127.65, 127.71, 127.82, 128.17, 128.21, 128.26, 137.79, 137.82, 138.22; IR (neat) 3063, 3030, 2926, 2857, 1607, 1497, 1454, 1360, 1308, 1260, 1208, 1105, 735, 698; MS (EI) m/z 91 (Bn, base peak), 532 (M⁺); HRMS (EI); calcd for C₃₄H₄₄O₅ 532.3189 found 532.3188.
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